# Hydrogen Sulfide: Neurophysiology and Neuropathology

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#### **Abstract**

Hydrogen sulfide (H<sub>2</sub>S) was known to be a toxic gas and an environmental hazard for many decades. However, it is now recognized that H<sub>2</sub>S may serve as a gaseous mediator that is endogenously produced to influence biological functions in mammalian. Together with nitric oxide and carbon monoxide, it forms the group of mediators that has been termed the "gasotransmitters." The past decade has seen an exponential growth of scientific interest in the physiological and pathological significance of H<sub>2</sub>S especially with respect to its role in the central nervous system and the cardiovascular system. In the central nervous system, H<sub>2</sub>S facilitates long-term potentiation and regulates intracellular calcium concentration and pH level in brain cells. Intriguingly, H<sub>2</sub>S produces antioxidant, anti-inflammatory, and antiapoptotic effects that may have relevance to neurodegenerative disorders. Abnormal generation and metabolism of H<sub>2</sub>S have been reported in the pathogenesis of ischemic stroke, Alzheimer's disease, Parkinson's disease, and recurrent febrile seizure. Exogenously applied H<sub>2</sub>S is demonstrated to have value for the treatment of febrile seizure and Parkinson's disease. This article presents an overview of current knowledge of H<sub>2</sub>S in relation to brain functions, with a special emphasis on its neuroprotective effects and the underlying cellular and molecular mechanisms. *Antioxid. Redox Signal.* 15, 405–419.

# Introduction

 $\mathbf{H}_{\text{COlorless}}$ , water-soluble gas with a characteristic smell of rotten eggs. At high concentrations (>500 ppm), H<sub>2</sub>S rapidly causes loss of consciousness and respiratory failure; thus, H<sub>2</sub>S poisoning is often associated with fatalities (58). Nevertheless, the notoriety of H<sub>2</sub>S as a toxic gas is experiencing a transformation, with increasing number of literature describing that it regulates a range of physiological and pathological processes in mammals. Thus, H<sub>2</sub>S is a physiologically important molecule and it has been referred to as the third gaseous mediator alongside nitric oxide (NO) and carbon monoxide (CO). H<sub>2</sub>S is largely produced from L-cysteine (Cys) and homocysteine (Hcy) by the actions of cystathionine  $\beta$ -synthase (CBS) and cystathionine  $\gamma$ -lyase (CSE). In general, CBS appears to be predominant in the central nervous system (CNS), whereas CSE is mainly expressed in the cardiovascular system (see the Biosynthesis and Metabolism of H<sub>2</sub>S in CNS section). H<sub>2</sub>S protects the heart against ischemia-reperfusion injury (17, 33) and elicits an antihypertensive effect by modulating vascular tone (26) and renin activity (50). It also induces a state of suspended animation in mouse (6), inhibits insulin release, and plays both pro- and anti-inflammatory roles in different systems and diseases [see (45) for review]. Further, H<sub>2</sub>S is revealed to be a novel neuromodulator (1). The actions of H<sub>2</sub>S in the CNS were first reported in 1996 by Abe and Kimura. These authors reported that  $H_2S$  at concentrations below  $130\,\mu M$  selectively enhances N-Methyl-D-aspartate (NMDA) receptor-mediated response and facilitates the induction of long-term potentiation (LTP), whereas at higher concentrations (>320  $\mu M$ ), it inhibits synaptic transmission in the hippocampus (1). Subsequently, more and more physiological and pathological functions of  $H_2S$  in the CNS were uncovered. In our previous reviews, we have focused on discussing the neurochemistry (56) and signaling properties of  $H_2S$  (65) in the CNS. In this article, we present current knowledge of  $H_2S$  to facilitate better understanding of its brain functions in both health and disease, with a special emphasis on its neuroprotective effects and the underlying cellular and molecular mechanisms involved.

# Chemical Properties of H<sub>2</sub>S

The solubility of  $H_2S$  in water ranges from 5.3~g/L at  $10^{\circ}C$  to 3.2~g/L at  $30^{\circ}C$  (54). It is weakly acidic because it dissociates into  $H^+$  and  $HS^-$  in solution. According to a standard Henderson-Hasselbach calculation, at  $20^{\circ}C$  and pH 7.4,  $H_2S$  exists as  $\sim 30\%$ –33%  $H_2S$  and 67%–70%  $HS^-$ , with negligible  $S^{2-}$  due to the high pK $_{a2}$  (>12). However, at  $37^{\circ}C$  and pH 7.4, <20% of  $H_2S$  exists as the undissociated form ( $H_2S$ ) (16). The term "free  $H_2S$ " is often used to refer to the sum of  $H_2S$ ,  $HS^-$ , and  $S^{2-}$ , as it is used in this article. Sometimes, the sum of  $H_2S$  and  $HS^-$  is denoted as total sulfide, but this may be

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misleading as this term may also be used for acid-labile sulfide and dithiothreitol-labile sulfide.

The alkali salts of  $H_2S$ , sodium hydrosulfide (NaHS), and sodium sulfide (Na<sub>2</sub>S) (or their hydrous forms) are widely used as exogenous sources of  $H_2S$  in laboratories. In aqueous solution, both can give a rapid bolus of  $H_2S$  at consistent concentrations, and thus their use is unlikely to reflect the biosynthesis and release of  $H_2S$  in vivo. To date, a small number of slow-releasing  $H_2S$  compounds such as GYY4137 and S-diclofenac are available and more novel compounds are under development (46, 47).

## Biosynthesis and Metabolism of H<sub>2</sub>S in CNS

The desulfhydration of Cys is proposed to be the major source of  $H_2S$  in mammals. This process is catalyzed via two pyridoxal-5'-phosphate (PLP)-dependent enzymes CBS and CSE. In the transsulfuration pathway, Cys is derived from Hcy with CBS catalyzing the  $\beta$ -replacement reaction of Hcy to yield cystathionine, which is then lyzed by CSE into Cys and  $\alpha$ -ketobutyrate (Fig. 1). CBS can efficiently produce  $H_2S$  via a  $\beta$ -replacement reaction in which Cys is condensed with Hcy to form cystathionine and  $H_2S$ , and this reaction is far more efficient when compared to  $\beta$ -elimination of Cys (9). Detailed kinetic analysis performed by Banerjee's group demonstrated that CBS produces  $H_2S$  overwhelmingly from

Cvs+Hcv (96%) under simulated physiological conditions, whereas Cys and Cys+Cys accounts for only 1%-3% (63). Therefore, cysteine and Hcy are the preferred substrates of CBS for H<sub>2</sub>S biosynthesis. On the other hand, CSE produces  $H_2S$  from Cys (70%) or Hcy ( $\gamma$ -elimination, 29%) under normal conditions (10  $\mu$ M Hcy). When the concentration of Hey was increased from 10 to 40 and 200  $\mu$ M to simulate mild and severe hyperhomocysteinemia, the contribution from Hcy increased from 29% to 63% and 90%, respectively, whereas contribution from Cys decreased correspondingly to 37% and 10%, respectively (63). As Vmax for the  $\gamma$ -elimination of Hcy is twice that for the  $\beta$ -elimination of Cys, this shift may represent a marked increase in the generation of H<sub>2</sub>S under hyperhomocysteinemic conditions. Therefore, H<sub>2</sub>S production derived from CSE is sensitive to Hcy (11). Basically, under normal conditions (10  $\mu$ M Hcy) CSE represents  $\sim 32\%$  of the H<sub>2</sub>S generation by the transsulfuration pathway, but it increases to  $\sim 45\%$  and  $\sim 74\%$  under moderate and severe hyperhomocysteinemia conditions (63). In contrast, CBS is not sensitive to Hcy concentrations with Cvs+Hcv as the predominant substrates. For this reason, in homocystinurics with CBS deficiency, CSE may be the major enzyme to produce H<sub>2</sub>S. Moreover, the level of H<sub>2</sub>S produced by CSE is predicted to be higher due to the enhanced accumulation of Hcy (11).

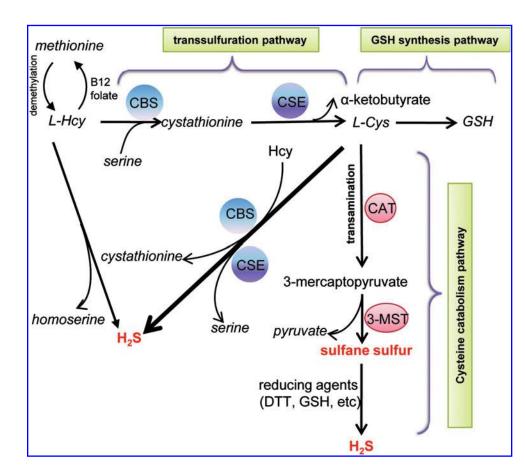


FIG. 1. Endogenous source of hydrogen sulfide (H2S) in mammalian. H<sub>2</sub>S is endogenously produced by the action of cystathionine  $\beta$ -synthase (CBS) and cystathionine  $\gamma$ lyase (CSE) in the transsulfuration pathway. By kinetic simulation, it is found that CBS generates H<sub>2</sub>S most efficiently from L-cysteine (Cys)+ homocysteine (Hcy), with cystathionine as a byproduct. reaction contributes > 95% of the net H<sub>2</sub>S production by CBS. On the contrary, the preferred substrates for CSE are Cys and Hcy. Together they contribute well over 90% of the net H2S production by CSE. In addition, the cysteine aminotransferase (CAT) and 3-mercaptopyruvate sulfurtransferase (3-MŠT) are components of the Cys catabolism pathway. CAT catalyzes the transamination of Cys to yield 3-mercaptopyruvate, a substrate of 3-MST to produce pyruvate and sulfane sulfur, which may liberate H<sub>2</sub>S in the presence of reductants such as dithiothreitol (DTT) and glutathione (GSH). The transsul-

furation pathway is critical for creating Cys from the essential amino acid methionine, which is first converted to Hcy by demethylation. CBS condenses serine and Hcy to produce cystathionine, which is converted to Cys and  $\alpha$ -ketobutyrate by CSE. The synthesis of GSH is regulated at the substrate level by Cys. Thus, the transsulfuration pathway also links to the GSH homeostasis in brain. (To see this illustration in color the reader is referred to the web version of this article at www.liebertonline.com/ars).

CBS is highly expressed in the brain and thus believed to be the primary physiologic source of H<sub>2</sub>S in the CNS (1), although both CBS and CSE activities were detected in different brain regions (3, 71). CBS is a cytoplasm PLP-dependent enzyme. Human CBS has a complex structure and regulatory mechanisms. It contains the N-terminal heme-binding domain, the catalytic domain, and the C-terminal regulatory domain. Two other gaseous transmitters, CO and NO, can bind to the heme-binding domain and result in the inhibition of CBS activity (67, 68). Moreover, the S-adenosyl-Lmethionine, which may bind to the C-terminal domain, can instantaneously activate CBS. At the transcriptional level, glucocorticoids can stimulate the CBS gene expression, whereas insulin can inhibit it (57). So far, at least 153 mutations in human CBS gene (see www.uchsc.edu/cbs/cbsdata/ cbsmain.htm) have been identified in patients with homocystinuria, a hereditary disease characterized by an accumulation of Hcy in the serum and uria. Several polymorphisms have been identified within the CBS gene. Apparently, there are gain-of-function polymorphisms (699C to T and 844\_ 845ins68) and loss-of-function polymorphism (1080C to T) (40). The two single-nucleotide polymorphisms in the CBS gene, 699C to T and 1080T to C, are associated with decreased risk of coronary artery disease and increased responsiveness to folic acid-induced total Hcy lowering effect.

The cellular localization of CBS is still controversial. Using immunohistochemical techniques, Robert et al. showed that CBS protein has a predominantly neuronal localization in most areas of the brain, especially in hippocampus and cerebellum (59). In contrast, Enokido et al. later demonstrated that CBS is preferentially expressed in astrocytes rather than neurons, which is verified by combined biochemical and histological examination, as well as in situ hybridization (18). This fits with recent findings that CBS mainly localizes to astrocytes (41). Lee et al. demonstrated that the basal H2S level in unstimulated human astrocytes is  $\sim 3.0 \,\mu\text{mol/g}$  protein, which is 7.9-fold higher than that in cultured microglia. More importantly, only astrocytes, instead of microglia, are strongly immunostained for CBS (41). However, Vitvitsky and his colleagues showed the incorporation of radiolabel from methionine into glutathione (GSH) in both cultured human astrocytes and neurons (71). Another group also showed that inhibition of CSE leads to a significant loss of GSH in adult brain slices (14). Since the only known route for the transfer of radiolabeled methionine to GSH is via the transsulfuration pathway involving CBS and CSE, these experiments indirectly justify the existence of CBS in both astrocytes and neurons. Nevertheless, studies consistently identified temporal expression of CBS in developing and adult mouse CNS. During the embryonic period, CBS protein level is generally low, but it dramatically increases from late prenatal to early postnatal period (18, 59).

In biomedical studies, small molecule inhibitors, such as hydroxylamine and aminooxyacetate acid (AOAA), have been used to determine the significance of the endogenously generated H<sub>2</sub>S. These agents are able to inhibit the biosynthesis of H<sub>2</sub>S from Cys, but they are general inhibitors of all PLP-dependent enzymes and are used quite to liberate bound PLP for quantitation (34, 64). In addition to heme and PLP, hydroxylamine also reacts with nonheme iron proteins, for example, ribonucleotide reductase and is used to inhibit cell growth. Hence, caution should be taken when interpreting results obtained from work involving these inhibitors.

In addition to CBS, there is also report showing that CSE plays an important role in human brain, despite its predominant localization in the cardiovasculature. In fact, CSE is critical for maintaining GSH homeostasis in brain, which in turn preserves mitochondrial function (14). CSE is the rate-limiting enzyme in the transsulfuration pathway for the sulfur transfer from methionine to Cys, which is a limiting reagent in the synthesis of GSH. Moreover, CSE mRNA is localized in brain and found to be predominantly present in neurons by in situ hybridization. The CSE activity in mouse brain was as low as 1% of the hepatic activity. However, in human brain the activity was 100 times more than that in mouse brain. Further, an intact transsulfuration pathway in the brain mediated by both CBS and CSE links to GSH homeostasis, which greatly contributes to the redox-buffering capacity in brain (71). Nevertheless, the general consensus is that CSE is the primarily physiological source of H<sub>2</sub>S generation in the peripheral tissues. There is definitive evidence that CSE knockout mice developed hypertension, which establishes that H<sub>2</sub>S is a major physiologic signaling molecule regulating vascular tone in mammals. So far, there is little knowledge about the physiologic relevance of CSE relative to CBS in brain, in addition to its role in transsulfuration pathway linking to GSH homeostasis. This issue merits further investigation.

Recently, Kimura's group reported another source of H<sub>2</sub>S in the brain homogenates of CBS-knockout mice (62). They show 3-mercaptopyruvate sulfurtransferase (3-MST) in combination with Cys aminotransferase (CAT) produces H<sub>2</sub>S from Cys. Like CBS and CSE, CAT is also a PLP-dependent enzyme that catalyzes the metabolism of Cys and  $\alpha$ -ketoglutarate to yield 3-mercaptopyruvate as the substrate for 3-MST. 3-MST is localized to mitochondria and nerve endings. As its name implies, it belongs to the family of sulfurtransferases, which catalyze the transfer of sulfane sulfur from persulfide or thiosulfate or mercaptopyruvate to an acceptor, and liberates H<sub>2</sub>S under certain conditions. Thus, 3-MST does not produce H<sub>2</sub>S by itself. Instead, it produces sulfane sulfur (or bound sulfur), which, in the presence of reductants like dithiothrietol used in in vitro assays, liberates H<sub>2</sub>S (34). Bound sulfur may be a source of H<sub>2</sub>S in brain and it can immediately release H<sub>2</sub>S in response to physiologic stimulation (31). This may explain why H<sub>2</sub>S was not depleted in the brain homogenates of CBS knock mice. Presumably, H<sub>2</sub>S is derived from this pool of sulfane sulfur under reducing conditions. However, the physiological significance of H<sub>2</sub>S derived from this source is yet to be determined with 3-MST knockout mice or other techniques. With respect to development, 3-MST protein expression in the mouse brain is maintained from embryonic day 16 to postnatal day 14 (P14) but downregulated between P28 and P52, and then increased slightly thereafter up to P156 (62).

However, the contributions and differences of CBS and 3-MST with respect to H<sub>2</sub>S generation under physiological and pathological conditions are still not clearly understood. As these two enzymes have different cell-type-specific expression profiles in the brain, it is possible that they may have different functions in various pathophysiological situations. It may be speculated that CBS may relate closely to the antineuroinflammatory role of H<sub>2</sub>S, whereas 3-MST may contribute more to the antioxidant action due to their different cellular localization.

In addition to biosynthesis, there are two forms of sulfur stores in mammals, acid-labile sulfur and bound sulfane sulfur (31). The former store, mainly localized to the iron–sulfur center of enzymes in mitochondria, releases H<sub>2</sub>S under acidic conditions, whereas the latter store, primarily localized to the cytoplasm, releases H<sub>2</sub>S under reducing conditions. The physiological importance of H<sub>2</sub>S released by bound sulfur remains unclear. However, the general consensus is that acid-labile sulfur is not a source of H<sub>2</sub>S under physiological conditions.

 $H_2S$  is proposed to undergo various chemical reactions during its catabolism in mammals. These include oxidation to sulfate, methylation to methanethiol, and dimethyl sulfide as well as reaction with metallo- or disulfide-containing proteins such as hemoglobin. However, its metabolic fates in cells remain elusive. *In vitro* experiments found that  $H_2S$  decays rapidly with a short life of  $\sim 10\,\mathrm{min}$  in cell culture (28, 74). Moreover, the beneficial effect of  $H_2S$  is still there even if the NaHS is removed by wash away before the subsequent exposure to oxidative stress-inducing insults (49). It is likely that  $H_2S$  could be stored and immediately released in response to physiological stimulation (31).

### Concentrations of Free H<sub>2</sub>S in Brain

Three methods employed for H<sub>2</sub>S measurement in the brain are methylene blue colorimetric assay, polarographic H<sub>2</sub>S sensor, and gas/ion chromatography combined with electrochemical detection. The polarographic  $H_2S$  sensor method is used for real-time measurement of H<sub>2</sub>S production in biological samples (15). It was reported that H<sub>2</sub>S was produced rapidly by brain supernatants at  $\sim 10.6$  pmol/s/mg protein (15). The gas/ion chromatography with electrochemical detection has also been applied for measuring H<sub>2</sub>S levels in brain (22), whereas methylene blue assay is relatively less used because it uses strong acid, which may lead to artificially elevated value due to the release of H<sub>2</sub>S from acid-labile sulfur. Early in 1989, Warenycia et al. first reported that rat brain tissue contained relatively high levels of free H<sub>2</sub>S ( $\sim 54 \,\mu M$ ) (72), and subsequent research work from various groups also showed high concentrations of H<sub>2</sub>S in brain (ranging 50-160 µM) in a variety of mammalian species, including rat, bovine, mouse, and human (22, 60). These values are frequently cited in the literature. However, more recent estimates indicate that the concentration of H<sub>2</sub>S in brain or plasma may be much lower, which is in the nanomolar range. Ishigami et al. found that H2S in brain is undetectable using gas chromatography with a detection limit of  $9.2 \,\mu M$ , where powdered silver was employed to absorb H<sub>2</sub>S, and N<sub>2</sub> gas, thiourea, as well as  $H_2SO_4$  were applied to release  $H_2S$  (31). Furne *et al.* reported that free  $H_2S$  level in brain is  $\sim 14$  nM by gas chromatography (19). In that study, the brain tissue was rapidly homogenized in a gas-tight syringe, and the concentration of H<sub>2</sub>S initially present in tissue was calculated from the H<sub>2</sub>S concentration in the gas space over the homogenate.

There seems to be a general consensus that the earlier measurements of  $50\text{--}160\,\mu\text{M}$  were almost certain to be overestimates arising from unintended conversions. These may include free  $\text{H}_2\text{S}$ ,  $\text{HS}^-$ , protein-bound sulfide such as acidlabile sulfur and dithiothreitol-labile sulfide or even the total sulfide in the tissues. Strong acid used for tissue process may lead to the release of acid-labile sulfur from iron–sulfur cen-

ters. Moreover, strong base may lead to the liberation of  $H_2S$  in the presence of reductants, which is commonly applied in some protein assays. The determination of  $H_2S$  in biological samples is often influenced by a number of factors such as its instability, high volatility, great susceptibility to oxidation, and release of sulfide out of the commonly used reagent dithiothreitol. Therefore, a more reliable and validated method with high sensitivity at the nanomolar range will be of great importance for the determination of the actual value of  $H_2S$  level in the brain.

#### H<sub>2</sub>S as a Neuromodulator

H<sub>2</sub>S serves as a neuromodulator that potentiates or inhibits the transmission of nerve impulses in neurons. For example, it is known to regulate LTP in hippocampus, a synaptic model of learning and memory. It selectively stimulates NMDA receptor-mediated currents via activating adenylyl cyclase and the subsequent cyclic adenosine monophosphate (cAMP)/ protein kinase A (PKA) cascades and thus facilitates the induction of LTP in the presence of a weak titanic stimulation (1, 36). However, H<sub>2</sub>S alone did not induce LTP, implying that H<sub>2</sub>S merely modulates LTP in active synapses (1). H<sub>2</sub>S could also promote astrocytic glutamate uptake, which plays an important part in clearing excessive glutamate from synaptic clefts and maintaining normal neurotransmission between neurons. These observations indicate that H<sub>2</sub>S plays an important modulatory role in the CNS (Fig. 2). In addition, Kombian et al. found that H2S reversibly inhibited both fast and slow synaptic responses in dorsal raphe serotonergic neurons (39). In another study, NaHS was shown to upregulate expression of  $\gamma$ -aminobutyric acid (GABA)<sub>B</sub> receptor subunits 1 and 2, whereas hydroxylamine downregulated expression of GABA<sub>B</sub> receptor subunit 2, but not that of GABA<sub>B</sub> receptor subunit 1 (24). As GABA is the major inhibitory neurotransmitters, this may also imply that H<sub>2</sub>S is critical in maintaining the excitatory/inhibitory balance in neurotransmission and thus support the proposal that H<sub>2</sub>S serves as a novel neuromodulator. However, as discussed earlier, one should be cautious with such interpretation as hydroxylamine has actions other than inhibiting CBS (34, 64).

Intracellular calcium ([Ca<sup>2+</sup>]<sub>i</sub>) homeostasis plays an important role in regulating synaptic activity and plasticity, as well as signal transmission between neuron and glial cells. H<sub>2</sub>S has been found to increase [Ca<sup>2+</sup>]<sub>i</sub> in neurons, astrocytes, and microglia by increasing calcium influx through L- and Ttype calcium channels and NMDA receptors located in plasma membrane, and calcium release from  $[Ca^{2+}]_i$  stores (44, 53, 77), as represented in Figure 3. Besides, both PKA and phospholipase C/PKC pathways mediate the action of H<sub>2</sub>S on  $[Ca^{2+}]_i$  (44, 77). In view of the reciprocal interactions between glia and neuron, H<sub>2</sub>S may thereby regulate synaptic activity by modulating the activities of both neurons and glia (53). Thus, these findings consolidate the neuromodulatory action of H<sub>2</sub>S in the CNS. Primary cultured human astrocytes synthesize  $H_2S$  at a rate of 15  $\mu$ mol/g protein/h, which is 7.5-fold higher than that in microglial cells, where H<sub>2</sub>S exhibits antiinflammatory and neuroprotective effects (41). Moreover, inflammatory stimulations of microglia and astrocytes cause downregulation of CBS and H<sub>2</sub>S synthesis (41).

In addition, H<sub>2</sub>S regulates intracellular pH (pH<sub>i</sub>) in rat primary cultured microglia and astrocytes through modu-

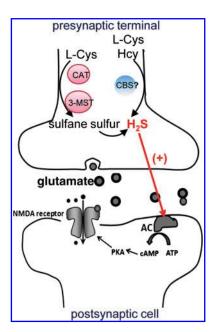


FIG. 2. Schematic diagrams showing the neuromodulatory role of H<sub>2</sub>S in central nervous system. The H<sub>2</sub>S produced by its generating enzyme, presumably CBS (its neuronal localization is still controversial), and the H<sub>2</sub>S liberated by sulfane sulfur under reducing conditions may activate adenylyl cyclase (AC) and its downstream protein kinase A (PKA) pathway to modulate N-Methyl-D-aspartate (NMDA) receptor activity and thus facilitates NMDA receptor-mediated long-term potentiation formation in the hippocampus. (To see this illustration in color the reader is referred to the web version of this article at www liebertonline.com/ars).

lating the activities of  $Cl^-/HCO_3^-$  exchanger and  $Na^+/H^+$  exchanger (Fig. 4) (48).  $pH_i$  homeostasis has an important role in the maintenance of normal cell function via changes in ion channel conductance, synaptic transmission as well as gap junctions.  $pH_i$  disturbance is an early event that occurs in brain under pathophysiological conditions such as hypoxia and ischemia. There is growing evidence demonstrating that the acid–base transporters contribute to the  $pH_i$  regulation in CNS. The regulatory effects of  $H_2S$  on  $pH_i$  via these transporters provide additional evidence that  $H_2S$  serves as a novel neuromodulator, not only under physiological conditions, but also in pathological situations.

### H<sub>2</sub>S as a Neuroprotectant

At micromolar range,  $H_2S$  has been demonstrated to show neuroprotective (antinecrotic and antiapoptotic) effects through multiple mechanisms in a series of *in vitro* studies. The following sections summarize and discuss the possible mechanisms underlying the neuroprotection offered by  $H_2S$ .

# Anti-inflammation

Neuroinflammation is a complex response to brain injury involving the activation of glia, release of inflammatory mediators within the brain, and recruitment of peripheral immune cells. Neuroinflammation has emerged to be a contributing factor intricately related to the cascade of events leading to neurodegeneration. Abundant evidence from postmortem and *in vivo* studies support that neuroin-

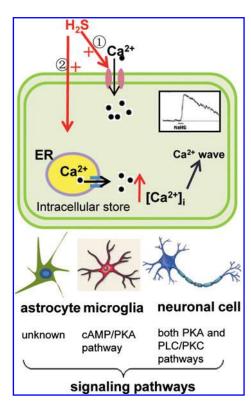


FIG. 3. Schematic diagrams showing the regulatory effects of  $H_2S$  on intracellular calcium ( $[Ca^{2+}]_i$ ) in both neuronal and glial cells and the underlying mechanisms involved.  $H_2S$  induces  $Ca^{2+}$  waves by increasing  $[Ca^{2+}]_i$  levels, which occurs via influx of extracellular  $Ca^{2+}$  through  $Ca^{2+}$  channels on cell membrane (labeled as ①) and release from  $[Ca^{2+}]_i$  stores (e.g., ER) (labeled as ②). Different signaling molecules are found to mediate these effects in different type of brain cells. For example, cAMP/PKA pathway contributes to the  $H_2S$  regulation of  $[Ca^{2+}]_i$  in rat microglia in primary culture. In SH-SY5Y neuron-like cells, both PKA and phospholipase C (PLC)/PKC pathways mediate the regulatory effects of  $H_2S$  on  $[Ca^{2+}]_i$  homeostasis. ER, endoplasmic reticulum; cAMP, cyclic adenosine monophosphate. (To see this illustration in color the reader is referred to the web version of this article at www.liebertonline.com/ars).

flammation is closely related to the pathogenesis of degenerative diseases such as Alzheimer's disease (AD) and Parkinson's disease (PD).

The activation of glial cells (especially microglia), which is a rapid cellular response to microenvironmental change, for example, neuronal injury, traumatic, or infectious stimuli, triggers deleterious events such as oxidative stress and cytokine-receptor-mediated apoptosis, leading to neuronal loss and possible disease progression. Hence, inhibition of the neuroinflammatory processes is a recognized therapeutic strategy aimed at delaying or halting the progression of neurodegenerative diseases. In 2007, our group first reported that NaHS attenuates lipopolysaccharide-induced production and release of NO and tumor necrosis factor alpha in primary cultured microglia and astrocytes, and murine immortalized BV2 microglial cells (29). In consistency, Lee et al. later demonstrated the antineuroinflammatory role of H<sub>2</sub>S and three H<sub>2</sub>S-releasing compounds (anethole trithione hydroxide, S-diclofenac, and S-aspirin) (42). H<sub>2</sub>S may exert antineuroinflammatory actions via inhibiting the production of

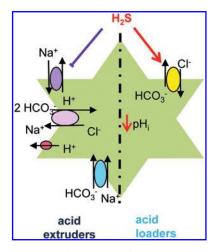


FIG. 4. Schematic illustration of the acid–base membrane transporters involved in intracellular pH regulation by H<sub>2</sub>S in glial cells. H<sub>2</sub>S decreases intracellular pH in glial cells *via* the stimulation of Na<sup>+</sup>-independent Cl<sup>-</sup>/HCO<sub>3</sub><sup>-</sup> exchanger and the inhibition of Na<sup>+</sup>/H<sup>+</sup> exchanger. (To see this illustration in color the reader is referred to the web version of this article at www.liebertonline.com/ars).

proinflammatory factors and enhancing the production of anti-inflammatory cytokines, as illustrated in Figure 5. Inhibition of p38/c-Jun-N-terminal kinase and nuclear factor kappa-light-chain-enhancer of activated B cells signaling pathways are recognized as possible mechanisms by which H<sub>2</sub>S restrains the extent of neuroinflammation and thereby limits the extent of neuronal injury. However, the actions of H<sub>2</sub>S on other inflammation-related intracellular molecules such as nuclear factor-erythroid 2-related factor 2, heat shock proteins, and matrix metalloproteinases still remain to be investigated. H<sub>2</sub>S may exert indirect neuroprotective effects via its anti-inflammatory role by which it inhibits proinflammatory factors released during microglial activation and thus alleviates neuroinflammation-related neurotoxicity (27, 42). Hu et al. found that the conditioned media from rotenone (10 nM)-treated microglia significantly decreased the cell viability of SH-SY5Y neuronal cells; however, this effect was alleviated in the neuronal cells treated with the conditioned media from NaHS plus rotenone cotreated microglia. At such low concentration, rotenone fails to decrease cell viability in SH-SY5Y cells, but it is enough to stimulate microglia activation. Hence, the observed protective effect of H<sub>2</sub>S, at least in part, arises from the suppression of proinflammatory factors released by rotenone-induced microglia.

# Antioxidation

Oxidative stress results from an overabundance of reactive free radicals secondary to either an overproduction of reactive oxygen species or a failure of cellular antioxidant buffering mechanisms such as GSH, a major and potent intracellular antioxidant. Excessive free radicals can react with essential molecules, including proteins, lipids, and nucleic acids and thereby disrupt their normal functions. Oxidative stress is implicated in the pathogenesis of neurodegenerative disorders, including AD and PD.  $\rm H_2S$  may act as a reducing agent. However, unlike the more abundant antioxidants (GSH present at  $1{\text -}10\,\mathrm{m}M$  concentration and Cys present at

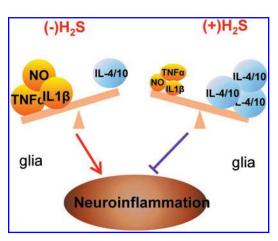
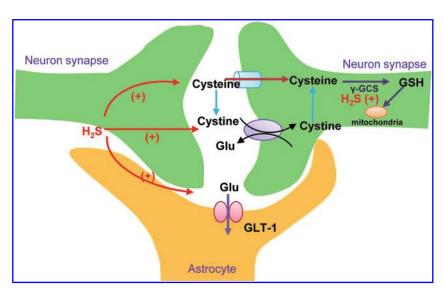


FIG. 5. Schematic diagrams illustrating the antineuroin-flammatory effects of  $H_2S$  in microglia and astrocytes. Microglia and, to a lesser extent, astrocytes mediate the neuroinflammatory processes in the central nervous system. These two types of glial cells, when activated appropriately, produce proinflammatory mediators that contribute to persistent neuroinflammatory responses. However, this process could be inhibited in the presence of  $H_2S$ , which may upregulate the production of anti-inflammatory cytokines such as interleukin (IL)-4/10 and downregulate the release of proinflammatory factors, including tumor necrosis factoralpha (TNF- $\alpha$ ), IL-1 $\beta$ , and nitric oxide (NO). (To see this illustration in color the reader is referred to the web version of this article at www.liebertonline.com/ars).

 $\sim 100 \,\mu M$  concentration), H<sub>2</sub>S is present at relatively low concentrations and it is also a poor reductant (redox potential of +0.17 vs. -0.25 V for the other two thiols) (34). Hence, the physiological relevance of the antioxidant properties by itself is an open question. However, NaHS, at the concentrations from 25 to 250  $\mu$ M, is able to enhance the reducing activity in neurons and protect them against oxidative damage induced by glutamate, hydrogen peroxide, or hypochlorous acid, primarily via increasing GSH levels instead of directly functioning as an antioxidant (38, 73). The GSH increase involves the stimulatory effects of  $H_2S$  on the activity of  $\gamma$ -glutamylcysteine synthetase, cystine, and Cys transport in neurons as well as glutamate uptake in astrocytes (37, 38, 49) (as shown in Fig. 6). Cys availability is a rate-limiting factor in GSH synthesis. Extracellular Cys is easily oxidized to cystine. The transport of cystine into cells, mainly mediated by a cystine/glutamate antiporter system Xc<sup>-</sup>, is therefore essential in providing cells with Cys as substrates for GSH synthesis. Excess extracellular glutamate may suppress cystine transportation into cells via Xc-. The excessive glutamate in the synaptic cleft can be cleared by neighboring glial cells through glutamate uptake via the excitatory amino acid transporters (EAATs). In astrocytes, glutamate uptake is mainly mediated by two EAATs subtypes: EAAT1 and EAAT2 (also known as GLAST and GLT-1, respectively). As GSH has a high turnover in cells, the inhibition of cystine transport by excessive glutamate may lead to rapid GSH depletion, which in turn increases the vulnerability of cells to oxidative injuries and ultimate cell death. Our group recently found that NaHS at 100 μM promotes [<sup>3</sup>H]glutamate uptake in astrocytes *via* enhancing the trafficking of glial glutamate transporter GLT-1 (49). This may not only lower extracellular glutamate and relieve the inhibition by glutamate on cystine transportation,

FIG. 6. Schematic paradigms illustrating the mechanisms for the elevation of GSH induced by H<sub>2</sub>S in brain cells. H<sub>2</sub>S enhances the transportation of cystine and Cys into cells to provide substrates for GSH synthesis in neurons. H<sub>2</sub>S also enhances glutamate uptake via glutamate transporter GLT-1 in astrocytes and thus clears the excessive glutamate in synaptic cleft. This process may also relieve the inhibition by glutamate on cystine transportation and thus facilitates the cystine transport into the neuronal cell. In addition, H<sub>2</sub>S enhances the activity of γ-glutamylcysteine synthetase ( $\gamma$ -GCS), a ratelimiting enzyme in GSH synthesis, and facilitates the redistribution of GSH into mitochondria and protects against oxidative stress. (To see this illustration in color the reader is referred to the web version of this article at www.liebertonline.com/ars).



but also yield the driving force for the cystine/glutamate antiporter  $Xc^-$  function that enables the transportation of cystine into cells, and eventually results in the elevation of intracellular Cys followed by an increase in intracellular GSH, and thus protects neurons against oxidative stress. These observations provide strong evidence for the powerful antioxidative action of  $H_2S$  in CNS, and also offer evidence for its neuroprotective effects because excitotoxicity, mainly derived from excessive accumulation of glutamate in the synaptic cleft, greatly contributes to the development of stroke, traumatic brain injury, and some neurodegenerative disorders.

# Antiapoptosis

Apoptosis, or programmed cell death, is a phenomenon that has been demonstrated to participate in neural development and play a role in neurodegeneration. Several pathological studies have revealed signs of apoptotic cell death in brains of PD and AD patients, although the extent is limited probably due to the slow progress of neurodegeneration. As the activation of apoptotic pathway most likely represents end-stage processes in neurodegeneration, the inhibitors of apoptosis have been proposed as potential neuroprotective agents regardless of the initial cause of neuronal loss. Mounting evidence shows that H<sub>2</sub>S has antiapoptotic effects on neuronal cells and thereby might become a potential candidate for neuroprotection. NaHS ( $<300 \,\mu M$ ) inhibits the apoptosis of PC12 and SH-SY5Y cells induced by various toxins, including 1-methyl-4-phenylpyridine, 6-hydroxydopamine (6-OHDA), rotenone, and  $\beta$  amyloid (66, 70, 76), all of which are commonly used in establishing in vivo and in vitro models for PD and AD. In addition, H<sub>2</sub>S protects hippocampal neurons against vascular dementia-induced injury via its antiapoptotic function (78). Most of the data reported so far indicate that the antiapoptotic effects of H<sub>2</sub>S mainly result from the preservation of mitochondrial integrity, that is, suppression of the mitochondrial apoptotic pathway (28, 76), as shown in Figure 7. H<sub>2</sub>S may inhibit the forming and opening of mitochondrial permeability transition pore and the subsequent release of cytochrome c from mitochondria to cytosol, as well as the activation of caspase cascades. These effects are dependent on the opening of mitochondrial ATP-

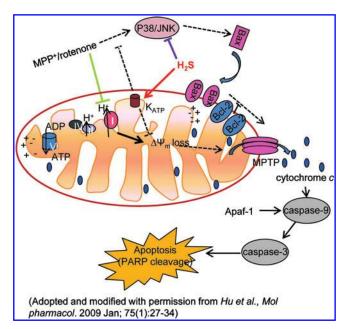


FIG. 7. Proposed signaling mechanisms for direct neuroprotection of H<sub>2</sub>S on neurotoxin-induced apoptosis. 1-Methyl-4-phenylpyridine (MPP<sup>+</sup>) or rotenone (complex I inhibitors) may initiate the  $\Delta\Psi_{m}$  dissipation and induce the mitochondrial permeability transition pore (MPTP) opening. These neurotoxins may also evoke the forming and opening of MPTP by activating p38/c-Jun-N-terminal kinase (JNK), inducing the release of cytochrome c from mitochondria to cytosol. The subsequently formed apoptosome (cytochrome c, apaf-1, and pro-caspase-9 complex) leads to the activation of caspase-9/3-dependent apoptotic pathway. The mechanisms underlying the anti-apoptotic effects of H<sub>2</sub>S may result from opening of mito-ATP-sensitive potassium channel (K<sub>ATP</sub>) channels, which in turn mediates the prevention of  $\Delta \Psi_{\rm m}$  loss and the inhibition of p38/JNK pathway. Adopted and modified with permission from Hu et al. (28). (To see this illustration in color the reader is referred to the web version of this article at www.liebertonline.com/ars).

sensitive potassium channels ( $K_{\rm ATP}$ ) (28). There is no report on the interaction between  $H_2S$  and death receptor apoptotic pathway. Brittain *et al.* recently identified the interaction of  $H_2S$  with human neuroglobin, a protein associated with mitochondria and protecting neurons from apoptotic stress (8); however, the biological significance of this interaction remains to be defined.

However, there is a study demonstrating that NaHS, at the concentration from 200 to  $1000 \,\mu M$ , greatly induced the mature cortical neurons apoptosis through the mechanisms involving the activation of calpain proteases and lysosomal destabilization, rather than caspases activation (10). Seemingly, this report is contradictory to the previous findings for the beneficial effects of H<sub>2</sub>S. However, NaHS, at a higher concentration up to  $1000 \,\mu\text{M}$ , yields  $\sim 333 \,\mu\text{M}$  H<sub>2</sub>S. This is much higher than the physiological range of  $H_2S$  (50–160  $\mu M$ ) in the brain, as discussed earlier. Such high levels of H<sub>2</sub>S may produce toxic effects on cells via inhibition of cytochrome c oxidase or other unidentified targets. In sum, several studies indicate that H<sub>2</sub>S exerts divergent effects on various brain cells and different animal disease models by using NaHS at various concentrations/doses. Some of them are summarized in Table 1. It is likely that H<sub>2</sub>S may play a neuroprotective role at physiological concentrations but may exhibit neurotoxic effects at significantly higher concentrations. In this sense, H<sub>2</sub>S shares similar features with another gaseous transmitter NO, which is also found to be neuroprotective at physiological amounts but clearly neurotoxic at higher concentrations. Additionally, similar to NO-induced protein S-nitrosylation, protein S-sulfhydration has recently been proposed to be a mechanism for H<sub>2</sub>S-mediated signaling (52). Nitrosylation appears to inhibit protein activity, whereas sulfhydration seems to enhance it. Sulfhydration is a post-translational modification. About 10%–25% of endogenous GAPDH,  $\beta$ tubulin, and actin are basally S-sulfhydrated and this sulfhydration directly enhances actin polymerization without affecting depolymerization (52). Of interest, the same group also showed that the stimulation of K<sub>ATP</sub> by H<sub>2</sub>S also arises from this protein-sulfhydrating effect (20). Therefore, protein S-sulfhydration may be an essentially molecular mechanism for H<sub>2</sub>S-mediated various biological effects.

All the aforementioned findings clearly indicate that H<sub>2</sub>S shows neuroprotective effects at low concentrations through various signaling pathways acting either cooperatively or independently. Then, how does H<sub>2</sub>S on earth exert these effects via a myriad of mechanisms? The answer seems to lie in the physical and chemical properties of this gaseous transmitter. H<sub>2</sub>S is a lipophilic molecule and it readily crosses cell membrane. This enables it to rapidly switch on or S-sulfhydrate intracellular targets. H<sub>2</sub>S not only modulates intracellular signaling molecules such as PKA and mitogen-activated protein kinases, but also regulates different ion channels such as L- and T-type calcium channel, K<sub>ATP</sub> channel, and chloride channel. H<sub>2</sub>S may react with these molecules within a few seconds, which in turn activate its downstream signaling pathways, resulting in the occurrence of biological functions of H<sub>2</sub>S in the CNS. However, no specific target/receptor has yet been identified presently. It is therefore proposed that H<sub>2</sub>S not only directly exerts effects in those cells where it is produced, but also acts on neighboring cells by diffusing into the surround and propagating the signal among neuron and glial cells, thus displaying a neuromodulatory role in the CNS.

#### H<sub>2</sub>S in CNS Diseases

Other than the physiological roles of a neuromodulator, there is increasing evidence that H<sub>2</sub>S is involved in the pathophysiology of CNS diseases such as epilepsy, stroke, Down's syndrome, AD, and PD, as summarized in Table 2. In addition, as mentioned above, the deficiency of CBS in humans results in homocystinuria, with increased plasma levels of Hcy and methionine but decreased levels of Cys. The clinical phenotype of these patients includes mental retardation, lens dislocation, and skeletal abnormality. This also implies the significance of H<sub>2</sub>S in health. Kinetic analysis has demonstrated that the CSE-catalyzed H<sub>2</sub>S production is sensitive to the level of Hcy and thus may increase in proportion to the grade of hyperhomocysteinemia (11). Hence, excessive production of H<sub>2</sub>S may be a contributing factor to the consequences of hyperhomocysteinemia. Remarkably, hyperhomocysteinemia is an independent risk factor for stroke (35) and has also been found to be correlated to a range of neurodegenerative disorders, including AD and PD. There is a significant increase of total Hcv level in the cerebrospinal fluid in patients with AD and PD (32). As discussed earlier, Hcy is maintained at relatively low concentrations and can be converted to Cys via the transsulfuration pathway. Cys is an essential substrate for the synthesis of GSH, the most abundant antioxidant in mammalian. In fact, oxidative stress and reduced GSH levels are common to the pathogenesis of neurodegenerative disorders. Therefore, the elevation of Hcy level and reduction of GSH level in AD and PD suggest the deficiency of CBS and/or CSE in the transsulfuration pathway (71), which links Hcy to GSH homeostasis. The recognition that CSE is sensitive to Hcy, whereas CBS is not should indicate an important role of CSE in relation to CNS diseases despite its predominant localization in the cardiovascular system. Further, in addition to inducing neuronal apoptosis, Hcy is also able to promote the proliferation and activation of microglia (80), which is a contributing factor to the pathological progress of neurodegenerative diseases. All in all, these findings indicate a correlationship between abnormal H<sub>2</sub>S biosynthesis and the development of the CNS diseases.

## Ischemic stroke

Ischemic stroke is caused by an interruption of blood supply to the brain (global) or part of the brain (focal) either by thrombosis or embolism. In a small clinical study, Wong et al. found that high plasma Cys level is correlated to poor clinical outcome 3 months poststroke in acute stroke patients (75). These authors further demonstrated that Cys loading by the intraperitoneal route dose dependently increases the infarct volume in rats after middle cerebral artery occlusion. Such effect of Cys was reversed by the coadministration of AOAA (a nonspecific inhibitor of PLP-dependent aminotransferases) (75) and mimicked by NaHS (55), indicating that H<sub>2</sub>S may be responsible for this effect. This is supported by the observation that the increased infarct caused by either Cys or NaHS was reversed by MK-801, an NMDA receptor blocker, as H<sub>2</sub>S is known to facilitate NMDA functions (36), thus enhancing the excitotoxicity triggered by ischemia. The dose at which NaHS mimicked the Cys loading effect was 0.18 mmol/kg, which is 66% of the reported  $LD_{50}$  for NaHS (72). In other words, H<sub>2</sub>S exacerbated tissue injuries in the rat middle cerebral artery occlusion model only at relatively high and most

TABLE 1. TOXIC AND PROTECTIVE EFFECTS OF HYDROGEN SULFIDE IN CELLS AND ANIMALS

	NaHS	Effects of H.C. Janeaus	Daf
	(μM)	Effects of H <sub>2</sub> S donors	Ref.
<i>In vitro</i> studies			
Rat hippocampal astrocytes	60–300	Increase [Ca <sup>2+</sup> ] <sub>i</sub> and induce calcium waves	53
Rat cortical microglia	100-500	Increase [Ca <sup>2+</sup> ] <sub>i</sub>	44
Rat cortical neurons	30–200	Increase intracellular GSH and protect against NMDA-induced excitotoxicity	38
SH-SY5Y neuroblastoma cells	25–250	Protect against hypochlorous acid-induced decrease in cell viability	73
	1–300	Inhibit rotenone-induced apoptosis via preservation of mitochondrial function	28
	1–50	NaHS and three other S-NSAIDs exert neuroprotective effects through their anti-inflammatory actions in microglia	42
PC12 cells	50–200	Protect against $A\beta_{(25-35)}$ -induced damage via scavenging ROS	66
	200-800	Inhibit MPP <sup>+</sup> -induced damage in PC12 cells	76
Mature mouse cortical neurons	200–1000	Induce neuronal apoptosis involves the activation of calpain proteases and lysosomal destabilization	10
Rat microglia and BV-2 cells	10–300	Suppress LPS-stimulated NO and TNF-α generation	29
<i>In vivo</i> studies	$(\mu \text{mol/kg})$	O	
Ischemic stroke	180 (i.p.)	Increase infarct size induced by ischemic stroke	75
Febrile seizure	56	Alleviate hippocampal damage induced by febrile seizure	25
Parkinson's disease	30, 100 (i.p.)	Exert neuroprotection in 6-OHDA and rotenone-induced model rat	27

 $[Ca^{2+}]_i$ , intracellular calcium; GSH, glutathione; H<sub>2</sub>S, hydrogen sulfide; LPS, lipopolysaccharide; MPP<sup>+</sup>, 1-methyl-4-phenylpyridine; NMDA, N-methyl-D-aspartate; NO, nitric oxide; 6-OHDA, 6-hydroxydopamine; ROS, reactive oxygen species; TNF- $\alpha$ , tumor necrosis factor-alpha. i.p., intraperitoneal injection; NaHS, sodium hydrosulfide; NSAIDs, non-steroid anti-inflammatory drugs.

certainly not physiological concentrations. Further, the endogenous levels of  $H_2S$  and its synthesizing activity in the affected cerebral cortex were significantly increased after middle cerebral artery occlusion (55).

More importantly, inhibition of  $H_2S$  production by various known inhibitors (AOAA and hydroxylamine,  $\beta$ -cyanoalanine, and DL-propargylglycine) preadministered before middle cerebral artery occlusion could reduce the infarct volume,

strongly indicating that increased H<sub>2</sub>S levels contributed significantly to the tissue infarction. Among these inhibitors, AOAA, at a dose of 0.05 mmol/kg, appears to be most effective followed by hydroxylamine which produces effect at 0.5–1.0 mmol/kg, in reducing middle cerebral artery occlusion-induced ischemic infarction. However, AOAA at higher doses (up to 0.5 mmol/kg) was not effective in ameliorating ischemic infarction (55). Seemingly, the inhibition of H<sub>2</sub>S synthesis

Table 2. Experimental Evidence for the Role of Hydrogen Sulfide in Central Nervous System Diseases

Disease	Evidence  Ischemic stroke increased tissue H <sub>2</sub> S in cerebral cortex.		
Ischemic stroke			
	Administration of cysteine and NaHS increases infarct size.	75	
AD	Total homocysteines are increased in AD brains and serum.	12	
	H <sub>2</sub> S attenuated beta-amyloid-induced damage in PC12 cells.	66	
	H <sub>2</sub> S attenuates LPS-induced cognitive impairment in rats.	21	
Febrile seizure	Increased plasma level of H <sub>2</sub> S and expression of CBS in hippocampus	25	
	of febrile seizure model rat NaHS improve hippocampal damage		
	induced by recurrent febrile seizure.		
HD	Plasma total homocysteine is increased in HD patients.	2	
	CBS interacts with huntingtin.	7	
PD	Plasma homocysteine levels are elevated in PD patients treated with L-dopa.	79	
	H <sub>2</sub> S exerts neuroprotective effects on neurotoxins-induced PD model rats.	27	

would be a potential approach for stroke therapy based upon these observations. However, it is premature to draw any conclusion for a number of still unresolved issues: (i) sublethal dose of NaHS administration was employed in this study; (ii) the inhibitors used are nonspecific inhibitors and may exhibit H<sub>2</sub>S-unrelated effects; (iii) the causality between stroke and endogenous H<sub>2</sub>S level elevation remains to be assessed in future. Moreover, in an *in vitro* study, NaHS (10–100  $\mu$ M) was shown to protect neurons against hypoxic injury via stimulation of K<sub>ATP</sub> channels (69), but at least, these studies strongly suggest that H<sub>2</sub>S is involved in the pathogenesis of ischemic stroke. It is highly likely that physiological levels of H<sub>2</sub>S exert a protective effect on cells against insults such as hypoxia. In the event of a stroke, overproduction of H<sub>2</sub>S may facilitate the cell death through enhancing excitotoxicity induced by excessive accumulation of extracellular glutamate. Whether or not inhibition of H<sub>2</sub>S production is a viable therapeutic approach for the treatment of acute stroke remains to be investigated. Nevertheless, elevated plasma Cys level in stroke patients could be used as an index to predict the poor outcome of stroke. Since H<sub>2</sub>S preconditioning-induced cardioprotection is well demonstrated, another important potential clinical application of H<sub>2</sub>S in stroke is in ischemic pre/postconditioning, which is deserving of attention in the future.

#### Alzheimer's disease

AD is the most common age-related neurodegenerative disorder. Its etiology remains unclear, but current evidence indicates the involvement of amyloid and tau proteins. In 1996, Morrison et al. first reported that the brain levels of Sadenosylmethionine, a CBS activator, are severely decreased in AD patients (51). The total serum level of Hcy (a precursor of Cys when acted on by CBS followed by CSE) is accumulated and increased in AD patients (12). One possible explanation is that the transsulfuration pathway linking Hcy and GSH metabolism, mediated by CBS and CSE, is disrupted. Because CBS is an important biosynthetic source of H<sub>2</sub>S generation in brain, although the contribution of 3-MST to H<sub>2</sub>S formation is also reported in brain, the dysfunction of the transsulfuration pathway may lead to the reduced production of H<sub>2</sub>S in AD, in addition to GSH. Indeed, several lines of evidence from both in vivo and in vitro studies indicate that H<sub>2</sub>S treatment elicits neuroprotective effects against pathological progression of AD. First of all, H<sub>2</sub>S is shown to scavenge the cytotoxic lipid oxidation product 4-hydroxynonenal (61), which is markedly increased in brains of severe AD patients. Second,  $H_2S$  was shown to ameliorate  $\beta$  amyloidinduced damage in PC12 cells through reducing the loss of mitochondrial membrane potential and attenuating the increase of intracellular reactive oxygen species (66). Third, H<sub>2</sub>S-releasing compounds are capable of attenuating neuroinflammation (42), a contributing factor implicated in AD pathogenesis. Importantly, H<sub>2</sub>S attenuates lipopolysaccharideinduced cognitive impairment in rats via its anti-inflammatory action (21). Additionally, garlic extracts, mainly the organosulfur-containing compounds such as S-allylcysteine and diallyl-disulfide, have been shown to reduce cerebral amyloid, inflammation, and tau conformational changes in AD transgenic model. Moreover, these garlic extracts (both fresh and boiled) not only inhibited  $\beta$  amyloid fibril formation but also was capable of defibrillating  $\beta$  amyloid preformed fibrils, thus exhibiting an antiamyloidogenic activity on amyloid-beta fibrillogenesis (23). H<sub>2</sub>S can be formed nonenzymatically from polysulfides in garlic (5). Based upon these findings, it is logical to assume that H<sub>2</sub>S would be beneficial for AD treatment. However, more direct evidence for the potential benefits of H<sub>2</sub>S or its donors in AD animal models is lacking at present.

#### Parkinson's disease

Like that in AD, plasma Hcy levels are also found to be elevated in PD patients treated with L-3,4-dihydroxyphenylalanine (79). However, little is known about the role of H<sub>2</sub>S in the initiation and development of PD. We recently demonstrated that H<sub>2</sub>S levels in the substantia nigra and striatum are considerably reduced in both 6-OHDA and rotenone-induced PD-like rats (27). More importantly, these authors found that systemic administration of NaHS (30 and 100 μmol/kg) dramatically attenuated the progression of movement dysfunction and loss of tyrosine hydroxylase positive-neurons in the substantia nigra induced by either rotenone or 6-OHDA. In addition, NaHS treatment inhibited the microglial activation in the substantia nigra and the accumulation of proinflammatory factors such as tumor necrosis factor-alpha and NO in the striatum. These observations indicate a role of endogenous H<sub>2</sub>S in the development of PD and thus its potential therapeutic value for PD treatment. This is supported by *in vitro* observations that H<sub>2</sub>S protects PC12 and SH-SY5Y cells against various neurotoxins (6-OHDA, 1methyl-4-phenylpyridine and rotenone) via antioxidative and antiapoptotic mechanisms (28, 70, 76). Besides the direct protective effects on neuronal cells, H<sub>2</sub>S is also able to indirectly protect SH-SY5Y cells against microglia-mediated neuroinflammatory toxicity induced by rotenone (27). This shows great disease-modifying significance because neuroinflammation is now recognized to be a critical factor in the pathogenesis of PD and other degenerative disorders. H<sub>2</sub>S exerts both direct and indirect neuroprotection on dopaminergic neurons, and thus modulating H<sub>2</sub>S synthetic pathways may become a potential approach for the treatment of PD (shown in Fig. 8). Hence, the recently developed H<sub>2</sub>Sreleasing L-3,4-dihydroxyphenylalanine hybrid molecules, which have been demonstrated to show antioxidant, antiinflammatory, and monoamine oxidase B inhibitory effects (43), would be of great value for PD treatment. However, the effects of H<sub>2</sub>S on the occurrence of Lewy's body, another important pathological feature in PD, have not been explored. Further, the physiological relevance of the H<sub>2</sub>S-generating enzymes such as CBS, CSE, CAT, as well as 3-MST in the development of PD is yet to be determined.

# Recurrent febrile seizure

Febrile seizure is the most common seizure type in children, often causing hippocampal damage. Han et~al. reported the plasma level of  $H_2S$  and expression of CBS in hippocampus were dramatically increased in the rat models of recurrent febrile seizure (25). In the same study, NaHS ( $56~\mu$ mol/kg) was found to lessen the hippocampal damage induced by recurrent febrile seizure, whereas hydroxylamine (12.5~mg/kg) aggravated this damage. The mechanisms by which NaHS alleviates the hippocampal damage involve its regulation of GABAB receptor function, as NaHS treatment could reverse downregulation of both GABAB receptor protein and mRNA in

the hippocampus of recurrent febrile seizure model rats (24). Therefore, H<sub>2</sub>S plays an important role in regulation of the GABAergic system besides its well-characterized actions on the glutamatergic system. It is thus not surprising that H<sub>2</sub>S may serve as a neuromodulator to maintain the excitatory/inhibitory neurotransmission in brain. Further, the increased H<sub>2</sub>S concentration as well as CBS expression in recurrent febrile seizure may be a compensatory response to suppress the neuronal hyperexcitability and thus alleviate the neuronal damage in hippocampus. Endogenous H<sub>2</sub>S may act in synergy with CO to protect against the hippocampal damage in recurrent febrile seizure because the researchers found that blockade of H<sub>2</sub>S production could reduce the CO level and the heme oxygenase expression, whereas administration of exogenous H<sub>2</sub>S could elevate them, and vice versa. Moreover, the different intracellular targets of these two gaseous transmitters [H<sub>2</sub>S acts on adenylyl cyclase (36), whereas CO affects guanylyl cyclase (13)] also point out the possible synergistic effect on memory processing. However, care should be taken in interpreting these observations as the inhibitor hydroxylamine used in that study can also permanently disable heme-containing molecules. Whether or not the interplay of these two gaseous molecules exerts a critical role in regulating the development of recurrent febrile seizure is yet to be defined.

#### Other CNS diseases

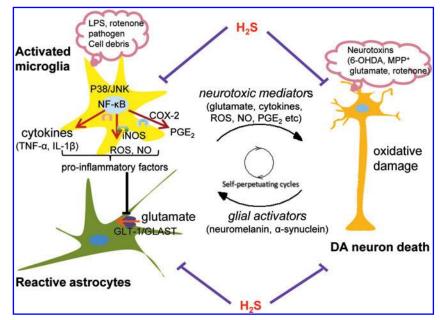
In addition to playing a part in the pathophysiology of aforementioned CNS diseases, H<sub>2</sub>S is also closely related to the pathogenesis of other CNS diseases such as Down syndrome and another degenerative disorder, Huntington's disease. Down syndrome is a chromosomal disorder characterized by the presence of all or part of an extra chromosome 21. A clinical study found that high level of thiosulfate—a catabolite of H<sub>2</sub>S, exists in the urine of Down syndrome patients, compared to that of the matched control subjects (4).

Further, clinical investigations have shown the overproduction of endogenous H<sub>2</sub>S in Down syndrome patients and thus established a correlationship between Down syndrome and chronic H<sub>2</sub>S poisoning. Excess H<sub>2</sub>S may be responsible for many clinical features of Down syndrome such as mental retardation. In fact, overproduction of H<sub>2</sub>S in Down syndrome is not surprising because the gene for CBS, one of the three enzymes in mammals responsible for H<sub>2</sub>S biosynthesis, is localized on chromosome 21 (21q22.3). In addition, Ichinohe et al. reported that CBS is enriched and localized to astrocytes and those surrounding senile plaques in the brains of Down's patients with AD (30). These observations imply that the overproduction of H<sub>2</sub>S, derived from CBS, may also be related to the formation and/or clearance of misfolded protein aggregates, which is a common pathological feature of neurodegenerative disorder. Compared to AD and PD, there is little knowledge about the role of H<sub>2</sub>S in Huntington's disease. There is only evidence that CBS interacts with huntingtin, mutants of which cause Huntington's disease. The plasma Hcy levels are also elevated in patients of Huntington's disease (2, 7). The elevation of plasma Hcy level in patients of the most common degenerative disorders, that is, AD, PD, and Huntington's disease, implicates a correlationship between Hcy aberration and the pathogenesis of neurodegenerative disorders. This field of research has been extensively carried out, but few are related to H<sub>2</sub>S aberration. Hence, further investigations are needed to determine the significance of H<sub>2</sub>S in this disorder.

## **Challenges and Limitations**

Although mounting evidence establishes the neuromodulatory function of  $H_2S$  in the CNS and thus suggests that modulating  $H_2S$  formation system may be a promising therapeutic approach for CNS diseases, researchers have come across challenges working with this gaseous molecule. First, the actual value of  $H_2S$  in brain tissues is still in

FIG. 8. Schematic paradigms illustrating the neuroprotective roles of H<sub>2</sub>S in Parkinson's disease. H<sub>2</sub>S shows direct protective effects on dopaminergic neurons against various neurotoxins (e.g., 6-hydroxydopamine [6-OHDA], MPP+ rotenone, and glutamate)-induced neuronal loss via antioxidant and antiapoptotic mechanisms. This may limit the injured dopaminergic neurons to release substances such as neuromelanin and αsynuclein, which are capable of activating glial cells in the injured site. In response to the stimuli resulting from external pathogen, neuronal injury, and/or neurotoxins, glial cells, especially microglia, are activated to release proinflammatory and neurotoxic factors such as TNF- $\alpha$ , IL-1 $\beta$ , NO, reactive oxygen species (ROS), and prostaglandin E<sub>2</sub> (PGE<sub>2</sub>), which may initiate or exacerbate dopaminergic neuronal damage, leading to the progression of Parkinson's disease. Moreover, H<sub>2</sub>S could enhance the glutamate uptake by astro-



cytes via GLT-1 and thereby reduces extracellular glutamate levels. Thus,  $H_2S$  may also exert indirect protection via its antineuroinflammatory role in microglia and astrocytes. (To see this illustration in color the reader is referred to the web version of this article at www.liebertonline.com/ars).

debate due to the methodological and technical problem of H<sub>2</sub>S determination. This affects the doses of H2S chosen for experimental application. The H<sub>2</sub>S levels achieved in the nigrostriatal tract is still unknown although it is reported that administration of NaHS at 1.68 and 5.6 mg/kg/day could be beneficial for rotenone/6-OHDA-induced PD model rats. Second, the therapeutic window for H<sub>2</sub>S treatment is quite narrow since lots of studies show that H<sub>2</sub>S produces beneficial effects at physiological concentrations (NaHS  $<300 \,\mu M$ ) but brings about harmful actions at significantly higher concentrations. Similar situation exists in the inflammation-regulatory effects by H<sub>2</sub>S. Li et al. points out that physiological concentrations of H<sub>2</sub>S produce anti-inflammatory effects, whereas higher concentrations, due to overaccumulation in already inflamed site, can exert proinflammatory effects (45). Third, several inhibitors of  $H_2S$  biosynthesis, such as AOAA and hydroxylamine, are commonly used for the exploration into the physiological relevance of endogenous H<sub>2</sub>S. Unfortunately, none of these are specific for H<sub>2</sub>S biosynthesis. In addition, few H<sub>2</sub>S donors except NaHS and Na<sub>2</sub>S are commercially available. Also, as mentioned earlier, no specific receptor/target of H<sub>2</sub>S has been defined. Last but not the least, although CBS<sup>-/-</sup> mice have been developed for testing the role of H<sub>2</sub>S in CBS predominantly expressed tissues especially in CNS, other sources of H<sub>2</sub>S, derived from CSE or released by bound sulfane under reducing conditions, could not be ignored. These may bring difficulties for the examination of the physiological significance of endogenous H<sub>2</sub>S in the CNS.

Further, attention should also be paid to the regimen of H<sub>2</sub>S given (bubbling, bolus, or slow-releasing of H<sub>2</sub>S) during experimentation. Bubbling of H<sub>2</sub>S gas gives a sustained source of H<sub>2</sub>S, but it also releases unpleasant smell in the working environment, which may bring about unhappiness to researchers and even animals. Moreover, NaHS gives a rapid bolus of H<sub>2</sub>S at a constant concentration. It also releases H<sub>2</sub>S more accurately and reproducibly than bubbling of  $H_2S\, gas\, in$ solution. Alternatively, the novel H<sub>2</sub>S-releasing compounds such as GYY4137 and S-diclofenac slowly release H<sub>2</sub>S and thus they may presumably reflect the enzymatic generation of H<sub>2</sub>S. For this reason, they would be more suitable for exploring the H<sub>2</sub>S biology in the CNS as well as other body systems. To address these abovementioned issues, more efforts should be put into the improvement for the techniques of H<sub>2</sub>S determination and the development of more commercially available H<sub>2</sub>S-releasing compounds. This will strengthen and enrich our knowledge of H<sub>2</sub>S biology in the CNS and other tissues.

### **Conclusions**

Accumulating evidence demonstrates that H<sub>2</sub>S confers pathophysiologically regulatory effects in brain, rather than being mere environmental toxin as previously described; therefore, now it must be considered as a biologically important molecule in both health and disease. This review summarizes the current knowledge on the effects of H<sub>2</sub>S in the CNS and discusses its roles in neuroprotection, as well as its therapeutic potential for neurodegenerative disorders. In summary, H<sub>2</sub>S acts as a neuromodulator in the CNS and may be involved in the pathogenesis of CNS diseases. Knowledge of H<sub>2</sub>S biology in the CNS highlighted here also raises the possibility of manipulating the H<sub>2</sub>S system for therapeutic

benefits to the patients suffering from AD, PD, and recurrent febrile seizure as well. However, in contrast to the great knowledge of the various biological functions of H<sub>2</sub>S in peripheral tissues, especially in the cardiovascular system, the exploration of H<sub>2</sub>S biology in the CNS is still at infancy. Great interest in unveiling this mystery is expected to be seen in the near future. Insight into the biological significance of H<sub>2</sub>S in brain may promote the understanding of the etiologies and pathologies of these diseases, perhaps leading to new treatment approaches. Bearing this in mind, intensive and extensive studies should be conducted in the near future to achieve a more comprehensive acknowledge of this gas in CNS, helping to seek promising and more effective therapeutic agents for neurodegenerative disorders (i.e., AD, PD, and Huntington's disease). However, due to the limitations mentioned above, there is still a long way to go before the mystery of H<sub>2</sub>S biology in the CNS could be uncovered and the application of any H<sub>2</sub>Sreleasing compound could be developed for clinical uses.

## **Acknowledgments**

This work was supported by Singapore research grants from National Medical Research Council (NMRC1183/2008 to J.-S.B.) and Biomedical Research Council (BMRC 08/1/21/19/559 to P.T.H.W.).

#### References

- Abe K and Kimura H. The possible role of hydrogen sulfide as an endogenous neuromodulator. *J Neurosci* 16: 1066–1071, 1996.
- 2. Andrich J, Saft C, Arz A, Schneider B, Agelink MW, Kraus PH, Kuhn W, and Muller T. Hyperhomocysteinaemia in treated patients with Huntington's disease homocysteine in HD. *Mov Disord* 19: 226–228, 2004.
- 3. Awata S, Nakayama K, Suzuki I, Sugahara K, and Kodama H. Changes in cystathionine gamma-lyase in various regions of rat brain during development. *Biochem Mol Biol Int* 35: 1331–1338, 1995.
- 4. Belardinelli MC, Chabli A, Chadefaux-Vekemans B, and Kamoun P. Urinary sulfur compounds in Down syndrome. *Clin Chem* 47: 1500–1501, 2001.
- Benavides GA, Squadrito GL, Mills RW, Patel HD, Isbell TS, Patel RP, Darley-Usmar VM, Doeller JE, and Kraus DW. Hydrogen sulfide mediates the vasoactivity of garlic. *Proc Natl Acad Sci U S A* 104: 17977–17982, 2007.
- Blackstone E, Morrison M, and Roth MB. H2S induces a suspended animation-like state in mice. Science 308: 518, 2005.
- Boutell JM, Wood JD, Harper PS, and Jones AL. Huntingtin interacts with cystathionine beta-synthase. *Hum Mol Genet* 7: 371–378, 1998.
- Brittain T, Yosaatmadja Y, and Henty K. The interaction of human neuroglobin with hydrogen sulphide. *IUBMB Life* 60: 135–138, 2008.
- 9. Chen X, Jhee KH, and Kruger WD. Production of the neuromodulator H2S by cystathionine beta-synthase via the condensation of cysteine and homocysteine. *J Biol Chem* 279: 52082–52086, 2004.
- Cheung NS, Peng ZF, Chen MJ, Moore PK, and Whiteman M. Hydrogen sulfide induced neuronal death occurs via glutamate receptor and is associated with calpain activation and lysosomal rupture in mouse primary cortical neurons. *Neuropharmacology* 53: 505–514, 2007.

- 11. Chiku T, Padovani D, Zhu W, Singh S, Vitvitsky V, and Banerjee R. H2S biogenesis by human cystathionine gammalyase leads to the novel sulfur metabolites lanthionine and homolanthionine and is responsive to the grade of hyperhomocysteinemia. *J Biol Chem* 284: 11601–11612, 2009.
- Clarke R, Smith AD, Jobst KA, Refsum H, Sutton L, and Ueland PM. Folate, vitamin B12, and serum total homocysteine levels in confirmed Alzheimer disease. *Arch Neurol* 55: 1449–1455, 1998.
- Cutajar MC and Edwards TM. Evidence for the role of endogenous carbon monoxide in memory processing. *J Cogn Neurosci* 19: 557–562, 2007.
- 14. Diwakar L and Ravindranath V. Inhibition of cystathionine-gamma-lyase leads to loss of glutathione and aggravation of mitochondrial dysfunction mediated by excitatory amino acid in the CNS. Neurochem Int 50: 418–426, 2007.
- Doeller JE, Isbell TS, Benavides G, Koenitzer J, Patel H, Patel RP, Lancaster JR Jr., Darley-Usmar VM, and Kraus DW. Polarographic measurement of hydrogen sulfide production and consumption by mammalian tissues. *Anal Biochem* 341: 40–51, 2005.
- Dombkowski RA, Russell MJ, and Olson KR. Hydrogen sulfide as an endogenous regulator of vascular smooth muscle tone in trout. Am J Physiol Regul Integr Comp Physiol 286: R678–R685, 2004.
- Elrod JW, Calvert JW, Morrison J, Doeller JE, Kraus DW, Tao L, Jiao X, Scalia R, Kiss L, Szabo C, Kimura H, Chow CW, and Lefer DJ. Hydrogen sulfide attenuates myocardial ischemia-reperfusion injury by preservation of mitochondrial function. *Proc Natl Acad Sci U S A* 104: 15560–15565, 2007
- 18. Enokido Y, Suzuki E, Iwasawa K, Namekata K, Okazawa H, and Kimura H. Cystathionine beta-synthase, a key enzyme for homocysteine metabolism, is preferentially expressed in the radial glia/astrocyte lineage of developing mouse CNS. *FASEB J* 19: 1854–1856, 2005.
- Furne J, Saeed A, and Levitt MD. Whole tissue hydrogen sulfide concentrations are orders of magnitude lower than presently accepted values. Am J Physiol Regul Integr Comp Physiol 295: R1479–R1485, 2008.
- 20. Gadalla MM and Snyder SH. Hydrogen sulfide as a gasotransmitter. *J Neurochem* 113: 14–26, 2010.
- Gong QH, Wang Q, Pan LL, Liu XH, Huang H, and Zhu YZ. Hydrogen sulfide attenuates lipopolysaccharide-induced cognitive impairment: a pro-inflammatory pathway in rats. *Pharmacol Biochem Behav* 96: 52–58.
- 22. Goodwin LR, Francom D, Dieken FP, Taylor JD, Warenycia MW, Reiffenstein RJ, and Dowling G. Determination of sulfide in brain tissue by gas dialysis/ion chromatography: postmortem studies and two case reports. *J Anal Toxicol* 13: 105–109, 1989.
- Gupta VB, Indi SS, and Rao KS. Garlic extract exhibits antiamyloidogenic activity on amyloid-beta fibrillogenesis: relevance to Alzheimer's disease. *Phytother Res* 23: 111–115, 2009.
- 24. Han Y, Qin J, Chang X, Yang Z, Bu D, and Du J. Modulating effect of hydrogen sulfide on gamma-aminobutyric acid B receptor in recurrent febrile seizures in rats. *Neurosci Res* 53: 216–219, 2005.
- 25. Han Y, Qin J, Chang X, Yang Z, Tang X, and Du J. Hydrogen sulfide may improve the hippocampal damage induced by recurrent febrile seizures in rats. *Biochem Biophys Res Commun* 327: 431–436, 2005.
- Hosoki R, Matsuki N, and Kimura H. The possible role of hydrogen sulfide as an endogenous smooth muscle relaxant

- in synergy with nitric oxide. Biochem Biophys Res Commun 237: 527–531, 1997.
- Hu LF, Lu M, Tiong CX, Dawe GS, Hu G, and Bian JS. Neuroprotective effects of hydrogen sulfide on Parkinson's disease rat models. *Aging Cell* 9: 135–146, 2010.
- 28. Hu LF, Lu M, Wu ZY, Wong PT, and Bian JS. Hydrogen sulfide inhibits rotenone-induced apoptosis via preservation of mitochondrial function. *Mol Pharmacol* 75: 27–34, 2009
- 29. Hu LF, Wong PT, Moore PK, and Bian JS. Hydrogen sulfide attenuates lipopolysaccharide-induced inflammation by inhibition of p38 mitogen-activated protein kinase in microglia. *J Neurochem* 100: 1121–1128, 2007.
- 30. Ichinohe A, Kanaumi T, Takashima S, Enokido Y, Nagai Y, and Kimura H. Cystathionine beta-synthase is enriched in the brains of Down's patients. *Biochem Biophys Res Commun* 338: 1547–1550, 2005.
- 31. Ishigami M, Hiraki K, Umemura K, Ogasawara Y, Ishii K, and Kimura H. A source of hydrogen sulfide and a mechanism of its release in the brain. *Antioxid Redox Signal* 11: 205–214, 2009.
- 32. Isobe C, Murata T, Sato C, and Terayama Y. Increase of total homocysteine concentration in cerebrospinal fluid in patients with Alzheimer's disease and Parkinson's disease. *Life Sci* 77: 1836–1843, 2005.
- 33. Johansen D, Ytrehus K, and Baxter GF. Exogenous hydrogen sulfide (H2S) protects against regional myocardial ischemia-reperfusion injury—evidence for a role of K ATP channels. *Basic Res Cardiol* 101: 53–60, 2006.
- 34. Kabil O and Banerjee R. Redox biochemistry of hydrogen sulfide. *J Biol Chem* 285: 21903–21907, 2010.
- 35. Kim NK, Choi BO, Jung WS, Choi YJ, and Choi KG. Hyperhomocysteinemia as an independent risk factor for silent brain infarction. *Neurology* 61: 1595–1599, 2003.
- 36. Kimura H. Hydrogen sulfide induces cyclic AMP and modulates the NMDA receptor. *Biochem Biophys Res Commun* 267: 129–133, 2000.
- Kimura Y, Goto Y, and Kimura H. Hydrogen sulfide increases glutathione production and suppresses oxidative stress in mitochondria. *Antioxid Redox Signal* 12: 1–13, 2010.
- 38. Kimura Y and Kimura H. Hydrogen sulfide protects neurons from oxidative stress. *FASEB J* 18: 1165–1167, 2004.
- Kombian SB, Reiffenstein RJ, and Colmers WF. The actions of hydrogen sulfide on dorsal raphe serotonergic neurons in vitro. J Neurophysiol 70: 81–96, 1993.
- 40. Kruger WD, Evans AA, Wang L, Malinow MR, Duell PB, Anderson PH, Block PC, Hess DL, Graf EE, and Upson B. Polymorphisms in the CBS gene associated with decreased risk of coronary artery disease and increased responsiveness to total homocysteine lowering by folic acid. *Mol Genet Metab* 70: 53–60, 2000.
- Lee M, Schwab C, Yu S, McGeer E, and McGeer PL. Astrocytes produce the antiinflammatory and neuroprotective agent hydrogen sulfide. *Neurobiol Aging* 30: 1523–1534, 2009.
- 42. Lee M, Sparatore A, Del Soldato P, McGeer E, and McGeer PL. Hydrogen sulfide-releasing NSAIDs attenuate neuroinflammation induced by microglial and astrocytic activation. *Glia* 58: 103–113, 2010.
- 43. Lee M, Tazzari V, Giustarini D, Rossi R, Sparatore A, Del Soldato P, McGeer E, and McGeer PL. Effects of hydrogen sulfide-releasing L-DOPA derivatives on glial activation: potential for treating Parkinson disease. *J Biol Chem* 285: 17318–17328, 2010.

44. Lee SW, Hu YS, Hu LF, Lu Q, Dawe GS, Moore PK, Wong PT, and Bian JS. Hydrogen sulphide regulates calcium homeostasis in microglial cells. *Glia* 54: 116–124, 2006.

- Li L, Bhatia M, and Moore PK. Hydrogen sulphide—a novel mediator of inflammation? Curr Opin Pharmacol 6: 125–129, 2006.
- Li L, Rossoni G, Sparatore A, Lee LC, Del Soldato P, and Moore PK. Anti-inflammatory and gastrointestinal effects of a novel diclofenac derivative. Free Radic Biol Med 42: 706– 719, 2007.
- 47. Li L, Whiteman M, Guan YY, Neo KL, Cheng Y, Lee SW, Zhao Y, Baskar R, Tan CH, and Moore PK. Characterization of a novel, water-soluble hydrogen sulfide-releasing molecule (GYY4137): new insights into the biology of hydrogen sulfide. *Circulation* 117: 2351–2360, 2008.
- 48. Lu M, Choo CH, Hu LF, Tan BH, Hu G, and Bian JS. Hydrogen sulfide regulates intracellular pH in rat primary cultured glia cells. *Neurosci Res* 66: 92–98, 2010.
- 49. Lu M, Hu LF, Hu G, and Bian JS. Hydrogen sulfide protects astrocytes against H(2)O(2)-induced neural injury via enhancing glutamate uptake. *Free Radic Biol Med* 45: 1705–1713, 2008.
- 50. Lu M, Liu YH, Goh HS, Wang JJ, Yong QC, Wang R, and Bian JS. Hydrogen sulfide inhibits plasma renin activity. *J Am Soc Nephrol* 21: 993–1002, 2010.
- 51. Morrison LD, Smith DD, and Kish SJ. Brain S-adenosylmethionine levels are severely decreased in Alzheimer's disease. *J Neurochem* 67: 1328–1331, 1996.
- 52. Mustafa AK, Gadalla MM, Sen N, Kim S, Mu W, Gazi SK, Barrow RK, Yang G, Wang R, and Snyder SH. H2S signals through protein S-sulfhydration. *Sci Signal* 2: ra72, 2009.
- 53. Nagai Y, Tsugane M, Oka J, and Kimura H. Hydrogen sulfide induces calcium waves in astrocytes. *FASEB J* 18: 557–559, 2004.
- 54. O'Neil MJ, Ann S, Heckelman PE, and Budavari S. (Eds). *Merck Index: An Encyclopedia of Chemicals, Drugs, and Biologicals*, 13th edition. Whitehouse Station, NJ: Merck and Co., Inc., 2001.
- 55. Qu K, Chen CP, Halliwell B, Moore PK, and Wong PT. Hydrogen sulfide is a mediator of cerebral ischemic damage. *Stroke* 37: 889–893, 2006.
- 56. Qu K, Lee SW, Bian JS, Low CM, and Wong PT. Hydrogen sulfide: neurochemistry and neurobiology. *Neurochem Int* 52: 155–165, 2008.
- 57. Ratnam S, Maclean KN, Jacobs RL, Brosnan ME, Kraus JP, and Brosnan JT. Hormonal regulation of cystathionine beta-synthase expression in liver. *J Biol Chem* 277: 42912–42918, 2002.
- Reiffenstein RJ, Hulbert WC, and Roth SH. Toxicology of hydrogen sulfide. *Annu Rev Pharmacol Toxicol* 32: 109–134, 1992.
- Robert K, Vialard F, Thiery E, Toyama K, Sinet PM, Janel N, and London J. Expression of the cystathionine beta synthase (CBS) gene during mouse development and immunolocalization in adult brain. *J Histochem Cytochem* 51: 363–371, 2003.
- 60. Savage JC and Gould DH. Determination of sulfide in brain tissue and rumen fluid by ion-interaction reversed-phase high-performance liquid chromatography. *J Chromatogr* 526: 540–545, 1990.
- Schreier SM, Muellner MK, Steinkellner H, Hermann M, Esterbauer H, Exner M, Gmeiner BM, Kapiotis S, and Laggner H. Hydrogen sulfide scavenges the cytotoxic lipid oxidation product 4-HNE. *Neurotox Res* 17: 249–256, 2010.

- 62. Shibuya N, Tanaka M, Yoshida M, Ogasawara Y, Togawa T, Ishii K, and Kimura H. 3-Mercaptopyruvate sulfurtransferase produces hydrogen sulfide and bound sulfane sulfur in the brain. *Antioxid Redox Signal* 11: 703–714, 2009
- 63. Singh S, Padovani D, Leslie RA, Chiku T, and Banerjee R. Relative contributions of cystathionine beta-synthase and gamma-cystathionase to H2S biogenesis via alternative transsulfuration reactions. *J Biol Chem* 284: 22457–22466, 2009.
- 64. Srivastava SK and Beutler E. A new fluorometric method for the determination of pyridoxal 5'-phosphate. *Biochim Biophys Acta* 304: 765–773, 1973.
- 65. Tan BH, Wong PT, and Bian JS. Hydrogen sulfide: a novel signaling molecule in the central nervous system. *Neurochem Int* 56: 3–10, 2010.
- 66. Tang XQ, Yang CT, Chen J, Yin WL, Tian SW, Hu B, Feng JQ, and Li YJ. Effect of hydrogen sulphide on beta-amyloid-induced damage in PC12 cells. Clin Exp Pharmacol Physiol 35: 180–186, 2008.
- 67. Taoka S and Banerjee R. Characterization of NO binding to human cystathionine beta-synthase: possible implications of the effects of CO and NO binding to the human enzyme. *J Inorg Biochem* 87: 245–251, 2001.
- 68. Taoka S, West M, and Banerjee R. Characterization of the heme and pyridoxal phosphate cofactors of human cystathionine beta-synthase reveals nonequivalent active sites. *Biochemistry* 38: 2738–2744, 1999.
- 69. Tay AS, Hu LF, Lu M, Wong PT, and Bian JS. Hydrogen sulfide protects neurons against hypoxic injury via stimulation of ATP-sensitive potassium channel/protein kinase C/extracellular signal-regulated kinase/heat shock protein90 pathway. *Neuroscience* 167: 277–286, 2010.
- Tiong CX, Lu M, and Bian JS. Protective effect of hydrogen sulfide against 6-OHDA induced cell injury in SH-SY5Y cells involves PKC/PI3K/Akt pathway. Br J Pharmacol 161: 467– 480, 2010.
- 71. Vitvitsky V, Thomas M, Ghorpade A, Gendelman HE, and Banerjee R. A functional transsulfuration pathway in the brain links to glutathione homeostasis. *J Biol Chem* 281: 35785–35793, 2006.
- 72. Warenycia MW, Goodwin LR, Benishin CG, Reiffenstein RJ, Francom DM, Taylor JD, and Dieken FP. Acute hydrogen sulfide poisoning. Demonstration of selective uptake of sulfide by the brainstem by measurement of brain sulfide levels. *Biochem Pharmacol* 38: 973–981, 1989.
- 73. Whiteman M, Cheung NS, Zhu YZ, Chu SH, Siau JL, Wong BS, Armstrong JS, and Moore PK. Hydrogen sulphide: a novel inhibitor of hypochlorous acid-mediated oxidative damage in the brain? *Biochem Biophys Res Commun* 326: 794–798, 2005.
- 74. Whitfield NL, Kreimier EL, Verdial FC, Skovgaard N, and Olson KR. Reappraisal of H2S/sulfide concentration in vertebrate blood and its potential significance in ischemic preconditioning and vascular signaling. *Am J Physiol Regul Integr Comp Physiol* 294: R1930–R1937, 2008.
- 75. Wong PT, Qu K, Chimon GN, Seah AB, Chang HM, Wong MC, Ng YK, Rumpel H, Halliwell B, and Chen CP. High plasma cyst(e)ine level may indicate poor clinical outcome in patients with acute stroke: possible involvement of hydrogen sulfide. *J Neuropathol Exp Neurol* 65: 109–115, 2006.
- Yin WL, He JQ, Hu B, Jiang ZS, and Tang XQ. Hydrogen sulfide inhibits MPP(+)-induced apoptosis in PC12 cells. *Life* Sci 85: 269–275, 2009.

- 77. Yong QC, Choo CH, Tan BH, Low CM, and Bian JS. Effect of hydrogen sulfide on intracellular calcium homeostasis in neuronal cells. *Neurochem Int* 56: 508–515, 2010.
- Zhang LM, Jiang CX, and Liu DW. Hydrogen sulfide attenuates neuronal injury induced by vascular dementia via inhibiting apoptosis in rats. *Neurochem Res* 34: 1984–1992, 2009.
- Zoccolella S, Lamberti P, Armenise E, de Mari M, Lamberti SV, Mastronardi R, Fraddosio A, Iliceto G, and Livrea P. Plasma homocysteine levels in Parkinson's disease: role of antiparkinsonian medications. *Parkinsonism Relat Disord* 11: 131–133, 2005.
- 80. Zou CG, Zhao YS, Gao SY, Li SD, Cao XZ, Zhang M, and Zhang KQ. Homocysteine promotes proliferation and activation of microglia. *Neurobiol Aging* 31: 2069–2079, 2010.

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Date of first submission to ARS Central, August 3, 2010; date of acceptance, September 2, 2010.

## **Abbreviations Used**

3-MST = 3-mercaptopyruvate sulfurtransferase

6-OHDA = 6-hydroxydopamine

AC = adenylyl cyclase

AD = Alzheimer's disease

AOAA = aminooxyacetate acid

[Ca<sup>2+</sup>]<sub>i</sub> = intracellular calcium

cAMP = cyclic adenosine monophosphate

CAT = cysteine aminotransferase

CBS = cystathionine  $\beta$ -synthase

CNS = central nervous system

CO = carbon monoxide

 $CSE = cystathionine \gamma$ -lyase

Cys = L-cysteine

DTT = dithiothrietol

EAATs = excitatory amino acid transporters

ER = endoplasmic reticulum

 $GABA = \gamma$ -aminobutyric acid

 $\gamma$ -GCS =  $\gamma$ -glutamylcysteine synthetase

GSH = glutathione

Hcy = homocysteine

HD = Huntington's disease

 $H_2S = hydrogen sulfide$ 

IL = interleukin

JNK = c-Jun-N-terminal kinase

 $K_{ATP} = ATP$ -sensitive potassium channel

LPS = lipopolysaccharide

LTP = long-term potentiation

 $MPP^+ = 1$ -methyl-4-phenylpyridine

MPTP = mitochondrial permeability transition pore

NaHS = sodium hydrosulfide

NMDA = N-Methyl-D-aspartate

NO = nitric oxide

PD = Parkinson's disease

 $PGE_2 = prostaglandin E_2$ 

pH<sub>i</sub> = intracellular pH

PKA = protein kinase A

PKC = protein kinase C

PLC = phospholipase C

PLP = pyridoxal-5'-phosphate

ROS = reactive oxygen species

TNF- $\alpha$  = tumor necrosis factor-alpha

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- 3. Jun Zhou, Peng-Fei Wu, Fang Wang, Jian-Guo Chen. 2012. Targeting gaseous molecules to protect against cerebral ischaemic injury: Mechanisms and prospects. *Clinical and Experimental Pharmacology and Physiology* **39**:6, 566-576. [CrossRef]
- 4. Reena V. Kartha, Jie Zhou, Laurie B. Hovde, Belinda W.Y. Cheung, Henning Schröder. 2012. Enhanced detection of hydrogen sulfide generated in cell culture using an agar trap method. *Analytical Biochemistry*. [CrossRef]
- 5. Leticia A. Montoya, Michael D. Pluth. 2012. Selective turn-on fluorescent probes for imaging hydrogen sulfide in living cells. *Chemical Communications* . [CrossRef]
- 6. Fengping Hou, Ju Cheng, Pinxian Xi, Fengjuan Chen, Liang Huang, Gouqiang Xie, Yanjun Shi, Hongyan Liu, Decheng Bai, Zhengzhi Zeng. 2012. Recognition of copper and hydrogen sulfide in vitro using a fluorescein derivative indicator. *Dalton Transactions*. [CrossRef]
- 7. Yong Qian, Ling Zhang, Shuting Ding, Xin Deng, Chuan He, Xi Emily Zheng, Hai-Liang Zhu, Jing Zhao. 2012. A fluorescent probe for rapid detection of hydrogen sulfide in blood plasma and brain tissues in mice. *Chemical Science* **3**:10, 2920. [CrossRef]
- 8. M. Ackermann, M. Kubitza, K. Maier, A. Brawanski, G. Hauska, A.L. Piña. 2011. The vertebrate homolog of sulfide-quinone reductase is expressed in mitochondria of neuronal tissues. *Neuroscience* **199**, 1-12. [CrossRef]
- 9. Eric R. DeLeon, Gilbrian F. Stoy, Kenneth R. Olson. 2011. Passive loss of hydrogen sulfide in biological experiments. Analytical Biochemistry . [CrossRef]
- 10. Ruma Banerjee . 2011. Hydrogen Sulfide: Redox Metabolism and Signaling. *Antioxidants & Redox Signaling* **15**:2, 339-341. [Abstract] [Full Text HTML] [Full Text PDF] [Full Text PDF] with Links]